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FCGR polymorphisms in the treatment of Rheumatoid Arthritis with Fc-containing TNF inhibitors

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Short title: FCGR gene polymorphisms in the response of RA to TNFi

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Abstract

We have explored if three FcyR functional polymorphisms could be biomarkers of the \bullet response of rheumatoid arthritis (RA) to the Fc-containing TNF inhibitors (TNFi) in samples from 429 patients. We also performed a meta-analysis of studies addressing FCGR3A F158V polymorphism, which is the most studied of the three. None of the three functional polymorphisms was associated with the response to TNFi. Meta-analysis of the seven FCGR3A F158V studies with available data, which included 899 patients with RA, showed no significant association (OR=1.11, 95% CI=0.8-1.5; P = 0.5). These results are notable given the large size of this study relative to others addressing these polymorphisms and the appearance given by previous studies of association of the FCGR3A F158V SNP.

Aims: We aimed to explore if the response of patients with rheumatoid arthritis (RA) **Keywords:** Rheumatoid arthritis, TNF inhibitors, biologics, genetics, biomarkers, Fc

receptor, FCGRT, infliximab, etanercept, adalimumab

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Introduction

Treatment of RA frequently involves the use of biologic drugs [1, 2]. The first biologics introduced in RA treatment were the TNF inhibitors (TNFi) and they still are the first choice for many patients. The most frequently used are infliximab (INX), etanercept (ETC) and adalimumab (ADM). The three include the Fc of human IgG1. INX and ADM include it as part of their monoclonal antibody molecule, whereas ETC includes the Fc as part of the fusion between sTNFR and IgG1. This situation is common among biologics because the Fc of IgG1 provides favorable pharmacokinetics, including a long half-life [3]. As a consequence, these three TNFi are influenced by the Fc gamma receptors (FcyR), which are pharmacogenetic modifiers with known functional polymorphisms [4-11]. These polymorphisms could explain part of the variable response of RA patients to TNFi treatments, with some patients very effectively responding whereas others, about a third, do not respond to the treatment. Variability that poses a major problem for the control of RA because about a third of the patients do not respond to the treatment [1, 2, 12, 13]. This lack of efficacy is largely-partly drug-specific and the same non-responder patient improves when switched to a different drug, other TNFi or a biologic directed against a different target. The causes of the variable response are still unknown, but there is a lot of interest in finding biomarkers to avoid the trial and error approach that has been followed up to now. These trials are very inefficient because knowing whether a treatment is effective or not for a given patient can take up to 6 months [14]. The FcyRs are a family of glycoprotein surface receptors, including the high affinity receptor FcyRI, and the low affinity receptors FcyRIIA, FcyRIIIA, FcyRIIC, FcyRIIIB and FcyRIIB [15]. All of them are expressed in immune cells. They have appeared by duplication and diversification and they still conserve a high degree of homology.

Reflecting their common origin, all the genes encoding the low affinity receptors are tightly placed on chromosome 1q23 (in the above-listed order). These receptors mediate, upon recognition of immune-complexes, the activation of innate and acquired immunity. The exception is $Fc\gamma RIIB$, which is inhibitory. In addition to the $Fc\gamma Rs$, there are other proteins binding the Fc. They are heterogeneous, expressed outside the immune system and known as Fc-receptor related proteins. The most notable is FcRn, which is encoded by the *FCGRT* gene [16-18]. It was originally described in the placenta, where it mediates the transfer of IgG from the mother to the fetus. Now, we know that FcRn expression is widespread and that its principal function is to maintain IgG prolonged half-life. It recovers the IgG that has been secreted in the urine and the gut. In addition, it also rescues IgG from intracellular lysosomal degradation, bringing it back to the circulation.

The Fc γ Rs have a less noticiable effect on IgG half-life than FcRn, but they can modify the efficacy or bioavailability of Fc-containing biologics also through their involvement in multiple immune pathways, including phagocytosis, antibodydependent cellular cytotoxicity and cellular signaling [15-18]. Therefore, the functional polymorphisms affecting the Fc γ Rs and FcRn are potential biomarkers associated with the patient variability in the response to TNFi. These possible biomarkers include non-synonymous SNPs affecting the protein sequence of Fc γ Rs and a variable number tandem repeat, VNTR, of 37 bp in the *FCGRT* promoter that has been associated with differences in FcRn expression and binding to IgG [19]. Recently, we have found an association between one of these polymorphisms, the *FCGR2A* H131R SNP, and the response to INX [20]. Other Fc γ R polymorphisms have also been reported as associated with the response to TNFi, most notably the *FCGR3A* F158V SNP [4, 7, 9, 10]. However, not all studies are concordant [5, 6, 8, 11], and no definitive validation of these biomarkers has yet been obtained. Here, we have examined two Fc γ R functional polymorphisms and the VNTR of *FCGR1* in the same 429 patients with RA in which we found association of the *FCGR2A* SNP [20], and we have also summarized the available evidence for the most studied of them, the *FCGR3A* F158V SNP, through meta-analysis of all available studies.

Material and methods

Patients. Biologic-naive patients with RA according to the 1987 revised American College of Rheumatology classification criteria were included [21]. They were recruited from six Spanish and two Greek Rheumatology Units during the course of a previous study [20]. They were treated with INX, ETC, or ADM between 2000 and 2010. The indication of treatment, the choice of drugs, and the control of disease evolution were performed with independence of this study, during standard care of the patients. Evaluations included the DAS28 and the EULAR criteria at the start of treatment and at 3, 6, and 12 months. The DAS28 is a composite index of RA activity including the number of tender joints and swollen joints (28 joints maximum), laboratory results and patient status assessment. The EULAR criteria divide patients into three classes based on change in DAS28 from baseline ($\Delta DAS28 = current$ DAS28 - baseline DAS28) and current DAS28 both measured at the time of evaluation: good responders are those with $\Delta DAS28 \ge 1.2$ and <u>current</u> $DAS28 \le 3.2$; non-responders are all patients with $\Delta DAS28 \le 0.6$ and those with $\Delta DAS28 > 0.6$ but \leq 1.2 and with <u>current DAS28 > 5.1</u>; all the remaining patients are moderate responders [22]. There was a total of 429 patients, 245 of them completed follow-up at 3, 6, and 12 months. Six of the 429 patients initially included in the study were

excluded because they showed a baseline DAS28 <3.2. Clinical characteristics are detailed in Table 1. All the patients provided blood samples for DNA extraction and their informed written consent to participate in the study. The study was approved by the local ethics committees and by the Comite Etico de Investigacion Clinica de Galicia (Santiago de Compostela, Spain).

Genotyping. The nsSNPs, rs1050501 in FCGR2B and rs396991 in FCGR3A were genotyped by PCR amplification followed by single-base extension with the SNaPshot Multiplex Kit (Applied Biosystems, Foster City, CA, USA). These SNPs are commonly referred to by their protein alleles with the following correspondences: rs1050501 T>C as Ile>Thre or I232T, and rs396991 T>G as Phe>Val or F158V. The VNTR of FcRn is difficult to amplify because of the high-GC content of the sequence. Its protocol is described in more detail: we used PCR amplification with enhancer PCR buffer [23] and a Touchdown PCR protocol [24]. Number of copies of the VNTR was determined by size in agarose gel electrophoresis. We obtained valid VNTR results from 97.0 % of our original discovery collection (295 patients) but no additional samples were genotyped given the problems and lack of difference observed. In addition, The VNTR genotypes of 44 samples were verified with a second PCR method in which 2'deoxyguanosine was substituted by 7-deaza-2'deoxyguanosine (7-deaza-dGTP) (Roche Diagnostics, Mannheim, Germany) following the protocol previously described [25]. Finally, samples with different genotypes of the three polymorphisms were sequenced to assess the accuracy of results. All primer sequences and details of the PCR protocols are provided (Supplementary Note 1 and Supplementary Table 1). Statistical analysis. The Statistica 7.0 (Statsoft, Tulsa OK) software was used

thoroughly. Allele frequencies, odds ratios (O.R) and their 95% confidence intervals

(95% CI) were calculated from 2x2 contingency tables. The HWE of the SNPs was analyzed with the Guo and Thompson exact method [26]. Pairwise D'and r² measures of linkage disequilibrium (LD) were obtained with Haploview [27]. Treatment outcomes were considered as $\Delta DAS28$, main outcome, or according to the EULAR criteria, secondary outcome [28]. A generalized linear model in the first case, and a logistic regression model in the second were fitted. In-both cases all analyses, genotypes were considered according with an additive genetic model, which is the model that best accounts for complex traits. EULAR categories were compared either as good + moderate responders versus non-responders, or as good responders versus non-responders (leaving moderate responders out of the analysis). The specific type of analysis is indicated in the text. We also tested for interaction involving FCGR2A and FCGR3A nsSNPs because it has been previously reported [29, 30]. Two approaches were followed. First, we grouped the diplotypes (two-gene genotypes) according to the presence of high affinity alleles (H131 in FCGR2A and 158V in FCGR3A). This classification resulted in a variable with four levels ranging from 0 for patients that were homozygotes for 131R and F158, to 4 high affinity alleles for patients that were homozygotes for H131 and 158V. This new variable was regressed against the treatment outcomes. The second approach was to analyze the interaction term in regression models including the two SNPs and the multiplicative interaction between them. All these analyses included three possible confounding factors as covariates: baseline DAS28, patient gender and the specific TNFi. In addition, some analyses were done with the center of recruitment and the year of treatment as additional covariates to account for heterogeneity in the treatment and evaluation of the patients. Baseline DAS28, the specific TNFi and the year of treatment were significantly associated with response to treatment. In addition, there were differences in response

to treatment between some centers of recruitment. Gender was included because it is a common covariate in most RA studies and some reports indicate that it is an important factor in the response to treatment, but no significant difference was observed in our study. Bonferroni correction by the number of FcγR polymorphisms we have studied (4 including ref. [20]) was applied for interpretation of the results. Power of our study for the main outcome was estimated with the GPower software [31].

We searched PubMed, the ISI Web of Science <u>using the following terms "Arthritis"</u> and "FCGR3A" and "Polymorphism" or "TNF" or "treatment", and the references of all the articles we have found for studies analyzing the role of *FCGR3A* F158V SNP in the response of RA patients to TNFi. <u>All the studies describing original studies</u> addressing genetic association of response to treatment with any of the three TNFi mentioned above (INX, ADM, ETC) in patients with rheumatoid arthritis were

selected. Information relative to the sample size, the outcome, the genetic model, and the percentages of responders and non-responders was manually extracted. One type of analysis was the most common to mostin -previous studies. It involved comparison of responders and non-responders following a FF genotype recessive model (FF *vs*. FV-VV). This model⁴ was selected due to the wide availability of the corresponding effect size for fixed-effects meta-analysis with the inverse variance weighting method as available in the *meta* library of the R project [32]. Fixed-effects meta-analysis was used because this model is generally more powerful than the random effect model. Heterogeneity of effect sizes was evaluated with the inconsistency parameter I² derived from the Cochran Q statistic [33]. A high, moderate and low level of inconsistency was attributed to levels of I² over 75%, 50% and 25%, respectively. Power of the meta-analysis was evaluated with the approach described by Hedges and Pigot [34].

Results

Patient characteristics and their response to TNFi. Six of the 429 patients initially included in the study showed a baseline DAS28 <3.2. They were excluded from analysis because this low disease activity makes evaluation unreliable. The remaining 423 selected RA-patients showed characteristics of severe RA (Table 1) with a high percentage of erosive arthritis (84%). In addition, high mean baseline DAS28 (5.9) and high mean baseline Health Assessment Questionnaire (HAQ) score (1.5) showed their active disease with moderate to severe disability before starting treatment with TNFi. This status was observed in spite of the previous treatment with a mean of 2.5 different Disease Modifying Anti-Rheumatic Drugs (DMARDs). These patients were followed during treatment with their first biologic drug, which specifically was one of the three most common TNFi, INX (60.8 %), ETC (20.8 %) or ADM (18.4 %). Most patients (94.6 %) received combined therapy with a DMARD accompanying the TNFi. This treatment resulted in improvement of DAS28 at all times of follow-up, but about 20 % of the patients were non-responders.

Analysis of *FCGR3A* and *FCGR2B* nsSNPs and of the VNTR in *FCGRT*. The two nsSNPs were successfully genotyped in 98.8 % of the 423 samples. The nsSNP in *FCGR2B* is located in a sequence that is identical to the sequence of *FCGR2C*, which is a pseudogene bearing the ancestral allele at the site homologous with the I232T polymorphism. This circumstance makes it impossible to distinguish IT heterozygotes from TT rare allele homozygotes. Therefore, a carrier analysis of the 232T allele was performed. In turn, the VNTR in *FCGRT* was difficult to amplify, but it was genotyped with a 97.0 % success rate in the first 295 samples (no additional samples were analyzed). Only the 2 (11.9 %) and 3 (87.2 %) repeats of the VNTR were common. The alleles of 4 and 5 repeats were observed, but in less than 1% of the patients and were excluded from further analysis. All the quality control filters were passed: genotypes were in HWE, replicated samples showed concordant genotypes, and the frequencies observed in our patients were similar to the previously reported in Europeans.

Association with the response to TNFi was analyzed considering $\Delta DAS28$ -as the main outcome and classification by the EULAR criteria-as a secondary outcome. None of these outcomes showed significant association with the nsSNP or with the VNTR genotypes at any of the three times analyzed (Table 2). The FCGR2B polymorphism was nominally associated with the response assessed by the EULAR criteria at 3 months of follow-up (P = 0.03). However, this association is not considered as significant because it did not pass Bonferroni correction by the number of tests, and it was only present when comparing responders (good + moderate responders) with non-responders. It was not present when comparing good responders with nonresponders (P = 0.08), or in analysis of the primary outcome, $\Delta DAS28$, which is more sensitive a continuous quantitative variable supporting more sensitive regression analysis than the EULAR criteria, a categorical variable [35]. The results were not modified by the inclusion of the center of recruitment and the year in which treatment with the TNFi was started as covariates (Supplementary Table 2). These analyses were done to account for possible heterogeneity in the treatment and evaluation of the patients.

Some previous studies have reported interactions between the *FCGR3A* F158V and the *FCGR2A* H131R nsSNPs [29, 30]. We explored this possibility taking advantage of our previous analysis of the *FCGR2A* nsSNP in the same RA patients considered

here [20]. The combined analysis showed that the two *FCGR2A/FCGR3A* nsSNPs were in weak LD (pairwise D' = 0.38, 95% CI = 0.26-0.48, and $r^2 = 0.074$) in our samples. However, there was not significant interaction between them (Supplementary Table 3) and the total number of the high affinity alleles (H131 at FCGR2A and 158V at FCGR3A) at the two nsSNPs was not associated with the response to TNFi (not shown).

Meta-analysis of studies addressing FCGR3A F158V association. We completed our analyses by summarizing the studies that have addressed the role of the FCGR3A F158V nsSNP in the response or RA to TNFi. We searched for articles and abstracts in the ISI Web of Science, in PubMed and in the bibliography of the manuscripts we were finding. Eight reports were found according to our inclusion criteria (Table 3). They were considered together with the current study. The nine studies are heterogeneous in sample size, in the TNFi used for treatment, in the outcomes and in the results. Four studies (including the current study) were of more than 280 patients each, whereas the other five studies were of less than 80 patients each. Together the five small studies add to fewer patients (254) than any one of the three large studies. Four of the five small studies showed a significant association with the response to TNFi, in contrast with none of the four large studies. This overrepresentation of significant association in the small studies was indicative of publication bias as shown by analyzing a funnel plot of the studies (Supplementary Figure 1) [36]. The four studies reporting a significant association showed an improved response in the FF homozygous patients. They included either the three TNFi or only INX, but no differences between the TNFi have been reported.

Combination of the results by meta-analysis could not include two of the large studies because of incomplete information [5] or a different pattern of genotype frequencies [11]. The other seven studies contained information allowing for the comparison of treatment responders and non-responders between patients with the FF genotype and patients with the FV-VV genotypes. Specifically, responders (good + moderate responders) and non-responders according to the EULAR criteria were available for 5 studies, whereas in two other studies responders and non-responders according to ACR20 were available. The two classifications were considered as equivalent for meta-analysis. This decision was supported by the fact that two associated studies had used the EULAR criteria [22], and the other two the ACR20 criteria [37]. There was also some heterogeneity in the times of assessment. However, the three months assessment was available in five studies and we choose this time point for evaluation. In the two studies lacking this time point, the nearest evaluation was considered. Two of the studies did not include patients that were both FF homozygotes and nonresponders [9, 10], making impossible to obtain an OR for them. These two studies were pooled with the most similar of the other studies to allow their inclusion in the meta-analysis. In this way, we analyzed 899 patients from seven studies (Figure 1). They did not show association with the response to TNFi (OR = 1.11, 95% CI = 0.8-1.5; P = 0.5). This result was observed with moderate heterogeneity between the studies ($I^2 = 62$ %). A priori power of a meta-analysis as the performed was sufficient (with $1-\beta = 0.8$, for $\alpha = 0.05$) to exclude an effect size equivalent to OR = 1.31. We also did not find association in a second meta-analysis. This meta-analysis combined results at different times, choosing the time of follow-up showing the strongest association in each study (Supplementary Figure 2). It was designed to offset possible losses of sensitivity due to the heterogeneity in the assessment.

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Discussion

We did not find significant association of any of the three functional polymorphisms analyzed, FCGR3A F158V, FCGR2B I232T, and the VNTR of FCGRT, with the response of RA patients to TNFi. These disappointing results were obtained in spite of the sample size of the study, the largest analyzing any of these polymorphisms in relation with the response to TNFi. In addition, the three polymorphisms were plausible biomarkers of the efficacy of Fc-containing TNFi. Two of them, FCGR3A F158V and FCGR2B I232T, have been previously associated with several disease phenotypes in patients with RA including response to TNFi [4-10, 38, 39]. The third, VNTR in FCGRT, has not been studied in this context, but its involvement was likely given the strong influence of FcRn on the IgG half-life [16-18]. Finally, the study was feasible as shown by the identification in the same RA patient samples of a reproducible association of response to INX and the FCGR2A H131R polymorphism [20] and by the *a priori* power of a study of this size to detect (with $1-\beta = 0.8$ power for $\alpha = 0.05$) effects of the FCGR3A polymorphism corresponding to a slope of 0.3 or larger, a slope of ≥ 0.46 for the *FCGR2B* SNP, and a slope ≥ 0.52 for the VNTR in FCGRT at 6 months of follow-up. Our results are more conclusive regarding the FCGR3A F158V nsSNP, because the availability of previous studies allowed combining them by meta-analysis. This meta-analysis rules out its value as a biomarker of the RA response to TNFi and suggests that previously reported FCGR3A F158V associations are likely ascribable to false positive results. Previous support for FCGR3A F158V as a biomarker of the response to TNFi in RA patients was apparently strong. However, our meta-analysis has shown its weakness. The distribution of the results of all the studies in function of their sample sizes (Supplementary Figure 1) is very indicative of publication bias, with the significant

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associations overrepresented among the small studies [36]. Therefore, it is likely that studies of small sample size not finding association have also been done, but remain unpublished. This bias gives the appearance of a notable fraction of studies with significant association, but this is misleading as shown by the summary statistic of our meta-analysis. However, a note of caution on this meta-analysis is required because it had not enough power to detect mild effects, and because previous studies were heterogeneous in the follow-up times and in the assessment of response to treatment. The FCGR3A F158V SNP has also been proposed as a biomarker of the response to other biologics. The most consistent results involve increased Antibody Dependent Cell Citotoxycity (ADCC) or killing by NK cells afforded by the V allele of FCGR3A F158V [40-42]. This mechanism seems to explain the improved response of some lymphomas and other hematological cancers to rituximab, an anti-CD20 chimeric monoclonal antibody, and other cancers to biologics that act by killing cancer cells [43]. It could also explain the better response to rituximab in patients with RA and other autoimmune diseases who carried the V allele of FCGR3A F158V [44-47]. In these diseases, rituximab efficacy is associated with the killing of autoimmune B cells. Therefore, the FCGR3A F158V SNP is a possible biomarker of the response to rituximab in RA, but we have ruled it out as a biomarker of the response to TNFi. This is in agreement with the lack of a role for ADCC and NK killing in the response to TNFi.

An additional complexity in the relationship of the *FCGR3A* F158V with Fccontaining drugs becomes evident when considering the studies analyzing Crohn's disease. In this disease, the VV genotype of *FCGR3A* F158V has been associated with the response to one of the TNFi, INX, but the improvement was observed in a biological response, not in the clinical response. In effect, several studies report a larger decrease of CRP in the VV homozygotes than in the other genotypes that implies some type of biological effect, but this was not translated in the clinical response [42, 48, 49]. We checked our patients for this effect, but no association between *FCGR3A* F158V and the decrease in CRP levels after treatment with INX was observed (Supplementary Table 4). All these results suggest that the *FCGR3A* F158V polymorphism influences the responses to treatment with some Fc-containing biologics with the clinical effect depending on the mode of action of the specific drug and on the specific disease.

The *FCGR2B* I232T SNP was not associated with the response to TNFi except for an isolated result observed at 3 months of follow-up with a particular comparison according to the EULAR criteria. This isolated observation obtained with our secondary outcome and not passing Bonferroni correction would most likely be due to chance assortment of the patients in the different response classes. However, lack of association with the response to TNFi should not be interpreted as a sign of the irrelevance of the polymorphism for other clinical phenotypes. In effect, there are sound studies showing association of the T allele with SLE susceptibility, especially in Asians [50].

The current study was conducted when four GWAS have already explored the response to TNFi in patients with RA [54-57]. These GWAS are small in relation with the studies analyzing disease susceptibility or other complex traits. The largest study, combining patients from the other 3 GWAS, included 2706 patients [57]. These numbers show the current difficulties in assembling large sets of patients with sufficient data for this type of genetic studies. Until now, the GWAS have identified some promising associations, as the association of CD84 with the response to ETC,

but none at the level of GWAS significance. None of these studies reported association with the $Fc\gamma R$ genes, which is <u>in</u> concordance with our results. However, this lack of reporting is hard to interpret because of the strict significance thresholds that GWAS apply.

In summary, none of the three functional polymorphisms in $Fc\gamma R$ genes explored here, the *FCGR3A* F158V and *FCGR2B* I232T nsSNPs and the VNTR in *FCGRT*, showed an association with the response to TNFi in patients with RA. These results were obtained in spite of the larger sample size relative to all the previous studies addressing these polymorphisms in the response to TNFi. In addition, meta-analysis of studies on the *FCGR3A* F158V SNP also showed lack of association. Therefore, the search of biomarkers to guide treatment of RA patients with TNFi should follow other routes.

Conflict of interest and sources of funding

There is not conflict of interest.

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Future prespectives

One of the main concerns in the management of RA patients is how to choose the most efficacious drug for a given patient given the individual variability in responses. This need is aggravated by the high cost of the biologics and the need to initiate an effective treatment early in the disease process to obtain the largest benefit for the patient. Already, there are several biomarkers for prediction of response to specific drugs that have been identified in sound studies, but none has yet been established as sufficiently reproducible or as clinically useful. Some of these biomarkers are pharmacogenetic biomarkers. They require further validation in additional sample collections as the evidence supporting them is not incontrovertible. This validation will be obtained by replication in new studies with large sample collections. In addition, they require an assessment of their predictive potential because studies until now have been centered in demonstrating a significant association not predictive power. This will require a different type of studies, most likely including clinical trials that incorporate the biomarkers in the assignement of patients to different treatment branches.

In addition, it is very likely that most genetic biomarkers are still undiscovered given the relative small size of the studies done until now. It is clear that this is the largest barrier for future progress: the lack of large collections of samples with response to treatment information. The recent availability of a variety of new drugs for treating RA does not make the tasks of collection these samples any easier, but the conscience of the problem and of the technical possibilities we have should convince more researchers to participate in these efforts. It is likely that after further study and validation the integration of different biomarkers from the genetics field and from other biological or clinical fields will lead to a clinical prediction model to help choose the most appropriate drug for each patient.

References

- 1. Nam JL, Winthrop KL, Van Vollenhoven RF *et al.*: Current evidence for the management of rheumatoid arthritis with biological disease-modifying antirheumatic drugs: a systematic literature review informing the EULAR recommendations for the management of RA. *Ann Rheum Dis* 69(6), 976-986 (2010).
- Smolen JS, Landewe R, Breedveld FC *et al.*: EULAR recommendations for the management of rheumatoid arthritis with synthetic and biological diseasemodifying antirheumatic drugs. *Ann Rheum Dis* 69(6), 964-975 (2010).
- 3. Lobo ED, Hansen RJ, Balthasar JP: Antibody pharmacokinetics and pharmacodynamics. *J Pharm Sci* 93(11), 2645-2668 (2004).
- Cañete JD, Suarez B, Hernandez MV *et al.*: Influence of variants of Fc gamma receptors IIA and IIIA on the American College of Rheumatology and European League Against Rheumatism responses to anti-tumour necrosis factor alpha therapy in rheumatoid arthritis. *Ann Rheum Dis* 68(10), 1547-1552 (2009).
- Criswell LA, Lum RF, Turner KN *et al.*: The influence of genetic variation in the HLA-DRB1 and LTA-TNF regions on the response to treatment of early rheumatoid arthritis with methotrexate or etanercept. *Arthritis Rheum* 50(9), 2750-2756 (2004).
- 6. Kastbom A, Bratt J, Ernestam S *et al.*: Fcgamma receptor type IIIA genotype and response to tumor necrosis factor alpha-blocking agents in patients with rheumatoid arthritis. *Arthritis Rheum* 56(2), 448-452 (2007).
- Morales-Lara MJ, Conesa-Zamora P, Garcia-Simon MS *et al.*: Association between the FCGR3A V158F polymorphism and the clinical response to infliximab in rheumatoid arthritis and spondyloarthritis patients. *Scand J Rheumatol* 39(6), 518-520 (2010).
- 8. Rooryck C, Barnetche T, Richez C, Laleye A, Arveiler B, Schaeverbeke T: Influence of FCGR3A-V212F and TNFRSF1B-M196R genotypes in patients with rheumatoid arthritis treated with infliximab therapy. *Clin Exp Rheumatol* 26(2), 340-342 (2008).
- 9. Tsukahara S, Ikari K, Sato E *et al.*: A polymorphism in the gene encoding the Fcgamma IIIA receptor is a possible genetic marker to predict the primary response to infliximab in Japanese patients with rheumatoid arthritis. *Ann Rheum Dis* 67(12), 1791-1792 (2008).
- 10. Tutuncu Z, Kavanaugh A, Zvaifler N, Corr M, Deutsch R, Boyle D: Fcgamma receptor type IIIA polymorphisms influence treatment outcomes in patients with inflammatory arthritis treated with tumor necrosis factor alpha-blocking agents. *Arthritis Rheum* 52(9), 2693-2696 (2005).
- 11. Sarsour K, Greenberg J, Johnston JA *et al.*: The role of the FcGRIIIa polymorphism in modifying the association between treatment and outcome in patients with rheumatoid arthritis treated with rituximab versus TNF-alpha antagonist therapies. *Clin Exp Rheumatol* 31(2), 189-194 (2013).
- 12. Malottki K, Barton P, Tsourapas A *et al.*: Adalimumab, etanercept, infliximab, rituximab and abatacept for the treatment of rheumatoid arthritis after the failure of a tumour necrosis factor inhibitor: a systematic review and economic evaluation. *Health Technol Assess* 15(14), 1-278 (2011).

- Salliot C, Finckh A, Katchamart W *et al.*: Indirect comparisons of the efficacy of biological antirheumatic agents in rheumatoid arthritis in patients with an inadequate response to conventional disease-modifying antirheumatic drugs or to an anti-tumour necrosis factor agent: a meta-analysis. *Ann Rheum Dis* 70(2), 266-271 (2011).
- 14. Emery P, Dorner T: Optimising treatment in rheumatoid arthritis: a review of potential biological markers of response. *Ann Rheum Dis* 70(12), 2063-2070 (2011).
- Nimmerjahn F, Ravetch JV: Fcgamma receptors: old friends and new family members. *Immunity* 24(1), 19-28 (2006).
- Ghetie V, Hubbard JG, Kim JK, Tsen MF, Lee Y, Ward ES: Abnormally short serum half-lives of IgG in beta 2-microglobulin-deficient mice. *Eur J Immunol* 26(3), 690-696 (1996).
- 17. Junghans RP, Anderson CL: The protection receptor for IgG catabolism is the beta2-microglobulin-containing neonatal intestinal transport receptor. *Proc Natl Acad Sci U S A* 93(11), 5512-5516 (1996).
- Zhu X, Meng G, Dickinson BL *et al.*: MHC class I-related neonatal Fc receptor for IgG is functionally expressed in monocytes, intestinal macrophages, and dendritic cells. *J Immunol* 166(5), 3266-3276 (2001).
- Sachs UJ, Socher I, Braeunlich CG, Kroll H, Bein G, Santoso S: A variable number of tandem repeats polymorphism influences the transcriptional activity of the neonatal Fc receptor alpha-chain promoter. *Immunology* 119(1), 83-89 (2006).
- 20. Montes A, Perez-Pampin E, Narvaez J *et al.*: Association of FCGR2A with the response to infliximab treatment of patients with rheumatoid arthritis. *Pharmacogenet Genomics* 24(5), 238-245 (2014).
- 21. Arnett FC, Edworthy SM, Bloch DA *et al.*: The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. *Arthritis Rheum* 31(3), 315-324 (1988).
- Van Gestel AM, Haagsma CJ, Van Riel PL: Validation of rheumatoid arthritis improvement criteria that include simplified joint counts. *Arthritis Rheum* 41(10), 1845-1850 (1998).
- 23. Ralser M, Querfurth R, Warnatz HJ, Lehrach H, Yaspo ML, Krobitsch S: An efficient and economic enhancer mix for PCR. *Biochemical and biophysical research communications* 347(3), 747-751 (2006).
- 24. Don RH, Cox PT, Wainwright BJ, Baker K, Mattick JS: 'Touchdown' PCR to circumvent spurious priming during gene amplification. *Nucleic Acids Res* 19(14), 4008 (1991).
- Musso M, Bocciardi R, Parodi S, Ravazzolo R, Ceccherini I: Betaine, dimethyl sulfoxide, and 7-deaza-dGTP, a powerful mixture for amplification of GC-rich DNA sequences. *The Journal of molecular diagnostics : JMD* 8(5), 544-550 (2006).
- 26. Guo SW, Thompson EA: Performing the exact test of Hardy-Weinberg proportion for multiple alleles. *Biometrics* 48(2), 361-372 (1992).
- 27. Barrett JC, Fry B, Maller J, Daly MJ: Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics* 21(2), 263-265 (2005).
- 28. Van Gestel AM, Prevoo ML, Van 'T Hof MA, Van Rijswijk MH, Van De Putte LB, Van Riel PL: Development and validation of the European League Against Rheumatism response criteria for rheumatoid arthritis. Comparison with the preliminary American College of Rheumatology and the World

Health Organization/International League Against Rheumatism Criteria. *Arthritis Rheum* 39(1), 34-40 (1996).

- 29. Niederer HA, Willcocks LC, Rayner TF *et al.*: Copy number, linkage disequilibrium and disease association in the FCGR locus. *Hum Mol Genet* 19(16), 3282-3294 (2010).
- Radstake TR, Petit E, Pierlot C, Van De Putte LB, Cornelis F, Barrera P: Role of Fcgamma receptors IIA, IIIA, and IIIB in susceptibility to rheumatoid arthritis. *J Rheumatol* 30(5), 926-933 (2003).
- 31. Faul F, Erdfelder E, Lang AG, Buchner A: G*Power 3: a flexible statistical power analysis program for the social, behavioral, and biomedical sciences. *Behav Res Methods* 39(2), 175-191 (2007).
- 32. Team RC: R: A Language and Environment for Statistical Computing. (2013).
- 33. Higgins JP, Thompson SG, Deeks JJ, Altman DG: Measuring inconsistency in meta-analyses. *Bmj* 327(7414), 557-560 (2003).
- Hedges LV, Pigott TD: The power of statistical tests in meta-analysis. *Psychol Methods* 6(3), 203-217 (2001).
- 35. Royston P, Altman DG, Sauerbrei W: Dichotomizing continuous predictors in multiple regression: a bad idea. *Stat Med* 25(1), 127-141 (2006).
- 36. Egger M, Davey Smith G, Schneider M, Minder C: Bias in meta-analysis detected by a simple, graphical test. *Bmj* 315(7109), 629-634 (1997).
- Felson DT, Anderson JJ, Boers M *et al.*: American College of Rheumatology. Preliminary definition of improvement in rheumatoid arthritis. *Arthritis Rheum* 38(6), 727-735 (1995).
- Chen JY, Wang CM, Ma CC *et al.*: A transmembrane polymorphism in FcgammaRIIb (FCGR2B) is associated with the production of anti-cyclic citrullinated peptide autoantibodies in Taiwanese RA. *Genes Immun* 9(8), 680-688 (2008).
- 39. Radstake TR, Franke B, Wenink MH *et al.*: The functional variant of the inhibitory Fcgamma receptor IIb (CD32B) is associated with the rate of radiologic joint damage and dendritic cell function in rheumatoid arthritis. *Arthritis Rheum* 54(12), 3828-3837 (2006).
- 40. Hatjiharissi E, Xu L, Santos DD *et al.*: Increased natural killer cell expression of CD16, augmented binding and ADCC activity to rituximab among individuals expressing the Fc {gamma}RIIIa-158 V/V and V/F polymorphism. *Blood* 110(7), 2561-2564 (2007).
- 41. Dall'ozzo S, Tartas S, Paintaud G *et al.*: Rituximab-dependent cytotoxicity by natural killer cells: influence of FCGR3A polymorphism on the concentration-effect relationship. *Cancer research* 64(13), 4664-4669 (2004).
- 42. Moroi R, Endo K, Kinouchi Y *et al.*: FCGR3A-158 polymorphism influences the biological response to infliximab in Crohn's disease through affecting the ADCC activity. *Immunogenetics* 65(4), 265-271 (2013).
- Mellor JD, Brown MP, Irving HR, Zalcberg JR, Dobrovic A: A critical review of the role of Fc gamma receptor polymorphisms in the response to monoclonal antibodies in cancer. *Journal of hematology & oncology* 6, 1 (2013).
- 44. Ruyssen-Witrand A, Rouanet S, Combe B *et al.*: Fcgamma receptor type IIIA polymorphism influences treatment outcomes in patients with rheumatoid arthritis treated with rituximab. *Ann Rheum Dis* 71(6), 875-877 (2012).

- 45. Kastbom A, Coster L, Arlestig L *et al.*: Influence of FCGR3A genotype on the therapeutic response to rituximab in rheumatoid arthritis: an observational cohort study. *BMJ open* 2(5), (2012).
- Robledo G, Marquez A, Davila-Fajardo CL *et al.*: Association of the FCGR3A-158F/V gene polymorphism with the response to rituximab treatment in Spanish systemic autoimmune disease patients. *DNA and cell biology* 31(12), 1671-1677 (2012).
- Quartuccio L, Fabris M, Pontarini E *et al.*: The 158VV Fcgamma receptor 3A genotype is associated with response to rituximab in rheumatoid arthritis: results of an Italian multicentre study. *Ann Rheum Dis* 73(4), 716-721 (2014).
- Louis E, El Ghoul Z, Vermeire S *et al.*: Association between polymorphism in IgG Fc receptor IIIa coding gene and biological response to infliximab in Crohn's disease. *Aliment Pharmacol Ther* 19(5), 511-519 (2004).
- 49. Louis EJ, Watier HE, Schreiber S *et al.*: Polymorphism in IgG Fc receptor gene FCGR3A and response to infliximab in Crohn's disease: a subanalysis of the ACCENT I study. *Pharmacogenet Genomics* 16(12), 911-914 (2006).
- Lee YH, Ji JD, Song GG: Fcgamma receptor IIB and IIIB polymorphisms and susceptibility to systemic lupus erythematosus and lupus nephritis: a metaanalysis. *Lupus* 18(8), 727-734 (2009).
- Gouilleux-Gruart V, Chapel H, Chevret S *et al.*: Efficiency of immunoglobulin G replacement therapy in common variable immunodeficiency: correlations with clinical phenotype and polymorphism of the neonatal Fc receptor. *Clin Exp Immunol* 171(2), 186-194 (2013).
- 52. Suzuki T, Ishii-Watabe A, Tada M *et al.*: Importance of neonatal FcR in regulating the serum half-life of therapeutic proteins containing the Fc domain of human IgG1: a comparative study of the affinity of monoclonal antibodies and Fc-fusion proteins to human neonatal FcR. *J Immunol* 184(4), 1968-1976 (2010).
- 53. Passot C, Azzopardi N, Renault S *et al.*: Influence of FCGRT gene polymorphisms on pharmacokinetics of therapeutic antibodies. *MAbs* 5(4), 614-619 (2013).
- Cui J, Stahl EA, Saevarsdottir S *et al.*: Genome-wide association study and gene expression analysis identifies CD84 as a predictor of response to etanercept therapy in rheumatoid arthritis. *PLoS Genet* 9(3), e1003394 (2013).
- 55. Liu C, Batliwalla F, Li W *et al.*: Genome-wide association scan identifies candidate polymorphisms associated with differential response to anti-TNF treatment in rheumatoid arthritis. *Mol Med* 14(9-10), 575-581 (2008).
- 56. Plant D, Bowes J, Potter C *et al.*: Genome-wide association study of genetic predictors of anti-tumor necrosis factor treatment efficacy in rheumatoid arthritis identifies associations with polymorphisms at seven loci. *Arthritis Rheum* 63(3), 645-653 (2011).
- 57. Umicevic Mirkov M, Cui J, Vermeulen SH *et al.*: Genome-wide association analysis of anti-TNF drug response in patients with rheumatoid arthritis. *Ann Rheum Dis* 72(8), 1375-1381 (2013).

Table 1.	Clinical	characteristics	of the	natients	with RA	included	l in	this	study
Table 1.	Cinnear	characteristics	or the	patients	with it's	monuace	111	uns	Study

Patients, number	<u>423</u>
Women, <u>%</u>	<u>83.9</u>
Age at diagnosis, median (IQR)	<u>45 (36-55)</u>
Diagnosis to anti-TNF, median years (IQR)	7 (3-12)
Rheumatoid factor, %	76.7
Anti-CCP antibodies, % *	70.2
Erosive arthritis, %	84.0
Smoking, %-*	16.4
$\overline{\text{DMARD before anti-TNF}, \text{ mean } \pm \text{SD}}$	2.5 ± 1.3
Concomitant DMARD, % ⁺	94.6
Baseline CRP (mg/L), median (IQR)*	8 (4-20)
Baseline ESR (mm/h), median (IQR)*	35 (19-56)
Baseline HAQ, median (IQR) ^a	1.5 (1-2.1)
TNF inhibitor, % (number)	
Infliximab (INX)	60.8 (257)
Adalimumab (ADM)	18.4 (78)
Etanercept (ETC)	20.8 (88)
DAS28, mean ± SD	
baseline	$\frac{5.9 \pm 1.2}{1.2}$
3 months	3.9 ± 1.4
6 months	3.8 ± 1.4
12 months	3.6 ± 1.4
EULAR response, %	
3 months	
responder	29.9
moderate	50.5
non-responder	19.6
6 months	
responder	39.9
moderate	39.7
non-responder	20.4
12 months	
responder	43.6
moderate	38.3
non-responder	18.1

Years from diagnosis to TNFi, median	7 (3-12)		
(IQR)	<u> </u>		
Rheumatoid factor, %	<u>76.7</u>		
Anti-CCP antibodies, % a	<u>70.2</u>		
Erosive arthritis, %	<u>84.0</u>		
Smoking, % ^a	<u>16.4</u>		
DMARD before anti-TNF, mean \pm SD	2.5 ± 1.3		
Concomitant DMARD, % a	<u>94.6</u>		
Baseline CRP (mg/L), median (IQR) a	<u>8 (4-20)</u>		
Baseline ESR (mm/h), median (IQR) a	<u>35 (19-56)</u>		
Baseline HAQ, median (IQR) ^a	<u>1.5 (1-2.1)</u>		
<u>TNF inhibitor, %</u>	INX / ADM / ETC ^b		
	<u>60.8 / 18.4 / 20.8</u>		
$DAS28$, mean \pm SD			
baseline	5.9 ± 1.2		
3 months	3.9 ± 1.4		
6 months	3.8 ± 1.4		
12 months	3.6 ± 1.4		
EULAR response, %	<u>R / M / NR^c</u>		
<u>3 months</u>	<u>29.9 / 50.5 / 19.6</u>		
<u>6 months</u>	39.9 / 39.7 / 20.4		
<u>12 months</u>	43.6 / 38.3 / 18.1		
85 % of the natients were available: 349 for a	nti-CCP antibodies 31	" Data from <	
261 for CRP 304 for ESR and 329 for baselin	ne HAO	i ioi sillokilig,	
$^{\text{b}}$ INX = infliximab, ADM = adalimumab. ET	C = etanercept		Formatted: Spanish (Spain)
$^{\circ}R = responder, M = moderate responder, NR$	= non-responder		Formatted: English (United States)

	FCGR3A F158V				FCGR2	B 1232T ^a			FCI	RG <i>T</i> ⁵			
	FF	FV	VV	_		П	IT+TT	_		3/2	3/3		
Total	35.6	51.4	12.9	-		74.6	25.4			19.3	80.7		
$\Delta DAS28$ (mean ± SD)				β	Ρ			β	Ρ			β	Р
3 months <u>, n = 364</u>	1.9 ±1.5	2.0 ±1.4	2.0 ±1.3	0.05	0.2	1.9 ±1.5	2.1 ±1.3	0.05	0.3	2.2 ±1.5	2.1 ±1.4	-0.07	0.3
6 months <u>, n = 377</u>	2.1 ±1.6	2.0 ±1.6	2.4 ±1.6	0.07	0.1	2.1 ±1.6	2.2 ±1.6	0.03	0.5	1.9 ±1.6	2.2 ±1.5	0.03	0.5
12 months <u>, n = 284</u>	2.4 ±1.6	2.2 ±1.8	3.0 ±1.7	0.06	0.2	2.3 ±1.7	2.4 ±1.7	0.001	1.0	2.2 ±1.8	2.5 ±1.7	0.02	0.7
EULAR criteria (n)				OR <u>(95%</u> <u>CI)</u>	Р			OR <u>(95%</u> <u>CI)</u>	Ρ			OR <u>(95%</u> <u>CI)</u>	Р
3 months <u>, n = 364 (364)</u>													
Responder	34.5	51.9	16.7	1.2	0.4	74.3	25.7	2.3	0.03	20.1	79.9	0.75	0.5
Non-responder	39.4	49.3	11.3	<u>(0.8-</u> <u>1.8)</u>		87.3	12.7	<u>(1.1-</u> 5.0)		17.4	82.6	<u>(0.3-</u> <u>1.8)</u>	
6 months <u>, n = 377 (377)</u>													
Responder	35.6	50.7	13.8	1.0	0.9	72.6	27.4	1.6	0.2	18.0	82.0	1.8	0.6
Non-responder	32.0	54.7	13.3	<u>(0.7-</u> <u>1.5)</u>		79.0	21.0	<u>(0.8-</u> <u>3.0)</u>		29.8	70.2	<u>(0.8-</u> <u>4.0)</u>	
12 months <u>, n = 284 (284)</u>													
Responder	34.5	52.6	12.9	1.4	0.2	72.5	27.5	1.03	0.9	16.8	83.2	1.4	0.5
Non-responder	32.7	63.5	3.8	<u>(0.8-</u> 2.3)		72.6	27.4	<u>(0.5-</u> 2.1)		22.2	77.8	<u>(0.5-</u> 3.6)	

Table 2. Distribution of the FCGR genotypes in the patients with RA.

Top row shows the genotype % for all patients. Upper rows present mean $\Delta DAS28 (\pm SD)$ for each genotype at the different evaluations. Lower rows show the genotype frequencies as percentage of the responder (good + moderate responders) and non-responder subgroups according to EULAR criteria. All analyses were adjusted by gender, TNFi and baseline DAS28

^a Only carrier analysis was possible for *FCGR2B*

^b The *FCGRT* VNTR was genotyped in all samples of our original discovery collection (295 patients) and exclude low frequency genotypes (2/2, 2/4, 3/4 and 3/5 that were observed only in 6, 2, 2 and 1 patients, respectively). Remaining patient numbers were: 245, 252 and 215 at 3, 6 and 12 months, respectively

 Table 3. Summary of the studies addressing association of the FCGR3A F158V

nSNP with the response to TNFi in patients with RA.

Study	Timeª	Number	Criteria	TNFi	% responders ^b	Association with response	
Sarsour <i>et al.</i>	6	390	CDAI℃	-	-	No	
Criswell <i>et al</i> .	12	301	ACR50	ETC	-	No	
Kastbom <i>et al.</i>	3	282	ACR20	INX	66.3	Νο	
	Ū	202		ETC	0010		
Rooryck <i>et al.</i>	3	78	ACR20	INX	56.4	No	
Tutuncu <i>et al.</i>	3	35	EULAR	INX ADM ETC	65.7	FF genotype	
Tsukahara <i>et al.</i>	5	29	EULAR	INX	93.1	FF genotype	
Morales-Lara <i>et al.</i>	3	38	ACR20	INX	76.3	No	
	6	37	EULAR		62.2	No	
	12	32			68.8	FF genotype (ACR	20)
Cañete <i>et al.</i>	1.5	73	ACR20	INX	67.5	FF genotype (ACR	20)
	7	74	EULAR	ETC ADM	61.0	No	
Current study	3	364	EULAR	INX ETC	80.5	No	
,	6	377		ADM	79.9	No	
	12	286			81.7	No	

^a Time of follow-up in months ^b Frequency of responders considering EULAR criteria (good + moderate responder) when available, ACR20 otherwise

^c CDAI = Clinical Disease Activity Index

Figure Legends

Figure 1. Fixed-effects meta-analysis of the studies comparing *FCGR3A* F158V between TNFi responder and non-responder patients with RA at 3 months of treatment. The Tutuncutu *et al.*, Morales-Lara *et al.* and Tsukahara *et al.* studies were joined to avoid zero counts in the OR denominator. EULAR response (good + moderate) was used when available; the ACR20 response otherwise. FF genotypes were compared with FV+VV genotypes.

Supplementary Material

It includes Supplementary Note 1 describing details of the protocols and Supplementary Table 1 with the primers and probes used for genotyping; Supplementary Table 2 with the association analysis after adjusting by center of recruitment and year of start of treatment as additional covariates; Supplementary Table 3 with the analysis of interaction between *FCGR2A* and *FCGR3A* nsSNPs; Supplementary Table 4 with analysis of the association between *FCGR3A* nsSNP and change in CRP at 6 months in patients treated with INX; and Supplementary Figure 1 with the funnel plot of all the studies addressing *FCGR3A* association with response to TNFi in RA patients; and Supplementary Figure 2 with the forest plot of a second meta-analysis including each study at the time of follow-up showing maximum association.

Executive summary

Need of biomarkers to select rheumatoid arthritis biologic drugs and opportunity among the *FCGR* genes

- A third of the patients with rheumatoid arthritis fail to respond to specific biologic drugs, but can respond after changing to another drug. The trial and error approach currently followed is inefficient and very costly.
- The pharmacokinetics and action of some of these biologic drugs (such as the TNF inhibitors: infliximab, etanercept and adalimumab) can be modulated by polymorphisms in the genes of FcyR because their molecules contain the Fc region of IgG1.

Study design and protocol

- Three functional FCGR polymorphisms were selected for study: FCGR3A F158V, which FF genotype has previously been associated with better response to TNF inhibitors in some studies, VNRT of FCGRT, which modulates the level of expression of the major determinant of IgG half-life, and FCGR2B I232T, which is associated with specific phenotypes in rheumatoid arthritis and other diseases
- Response to treatment was evaluated in 429 patients with RA treated with TNF inhibitors as the first biologic drug either as change in DAS28 or according to the EULAR response criteria.
- A meta-analysis of all published studies addressing the association of FCGR3A F158V with response to TNF inhibitors in rheumatoid arthritis was also performed.

Association of polymorphisms and clinical response to TNF inhibitors

- None of the three functional polymorphisms showed significant association with the response to TNF inhibitors.
- The meta-analysis of seven *FCGR3A* F158V with sufficient information showed also lack of significant association. This result contrast markedly with the previous appearance of association due to the small size of the four studies reporting association.

Interpretation

- None of the three FCGR polymorphisms studied influence response to TNF inhibitors in rheumatoid arthritis. However, other FCGR polymorphisms do modify this response as we have shown in a reproducible way for FCGR2A H131R.
- The apparent reproducibly association of *FCGR3A* FF genotype with a better response to TNF inhibitors can be ascribed to false positive results and publication bias as none of the larger studies, nor the meta-analysis supported this result.
- However, the *FCGR3A* F158V polymorphism can modify the clinical response to drugs with a different mechanism of action, as rituximab that cause cell cytotoxicity, or other types of responses as the modulation of CRP levels observed in Crohn's disease.

Conclusions

 None of the three functional FcγR polymorphisms explored here was associated with the response to TNFi in patients with RA.

Meta-analysis of all the FCGR3A F158V showed lack of association.